

ILLUSTRATED REVIEW

Protease-activated receptors: An illustrated review

Xu Han MS¹  | Marvin T. Nieman PhD¹   | Bryce A. Kerlin MD^{2,3}  

¹Department of Pharmacology, Case Western Reserve University, Cleveland, OH, USA

²Center for Clinical and Translational Research, Abigail Wexner Research Institute at Nationwide Children's Hospital, Columbus, OH, USA

³Department of Pediatrics, The Ohio State University College of Medicine, Columbus, OH, USA

Correspondence

Marvin T. Nieman, Department of Pharmacology, Case Western Reserve University, 2109 Adelbert Road W309B, Cleveland, OH, 44106-4965, USA.
Email: nieman@case.edu

Funding information

XH receives research funding from the American Heart Association Summer 2018 Predoctoral Fellowship (18PRE33960396) and cofunded by the Schwab Charitable Fund. MN receives research funding from the National Institutes of Health (HL098217). BAK receives research funding from the National Institutes of Health (K08DK103982 and R03DK118315) and the George & Elizabeth Kelly Foundation (Lewis Center, OH).

Handling Editor: Dr Alisa Wolberg

Abstract

Proteases are important regulators of cell behavior, survival, and apoptosis. They communicate to cells directly through a special class of G-protein-coupled receptors known as protease-activated receptors (PARs). N-terminal PAR proteolysis unmasks a neo-N-terminus, which serves as a tethered ligand to activate PARs. Using this unique irreversible activation mechanism, PARs relay information across cell membranes. The year 2020 is the 30th year since discovery of the first member of this family, PAR1. In this illustrated review, we highlight achievements in the PAR field over the past 3 decades. Additionally, the known expression profiles of PARs in human tissues and across species are portrayed. We also illustrate the tethered ligand activation mechanism, which is unique to PARs, and PAR regulatory mechanisms. PAR1 was originally named “thrombin receptor” because thrombin was the first protease identified to activate PAR1. However, over the past 30 years, a growing number of proteases have been found to cleave PARs and trigger differential downstream signaling depending on cleavage site, cell type, and species. We exemplify the diversity of PAR1-mediated signaling outcomes in platelets and endothelial cells as pertinent examples to the hemostasis, thrombosis, and vascular biology fields. Further, the termination and regulation of PAR signaling via endocytosis and currently available pharmacologic approaches are depicted. We conclude with portrayal of clinically translational aspects of PAR biology including pharmacologic manipulation and single-nucleotide polymorphisms.

KEYWORDS

antithrombotic therapies, G-protein-coupled receptors, platelets, protease-activated receptors, signaling, thrombosis

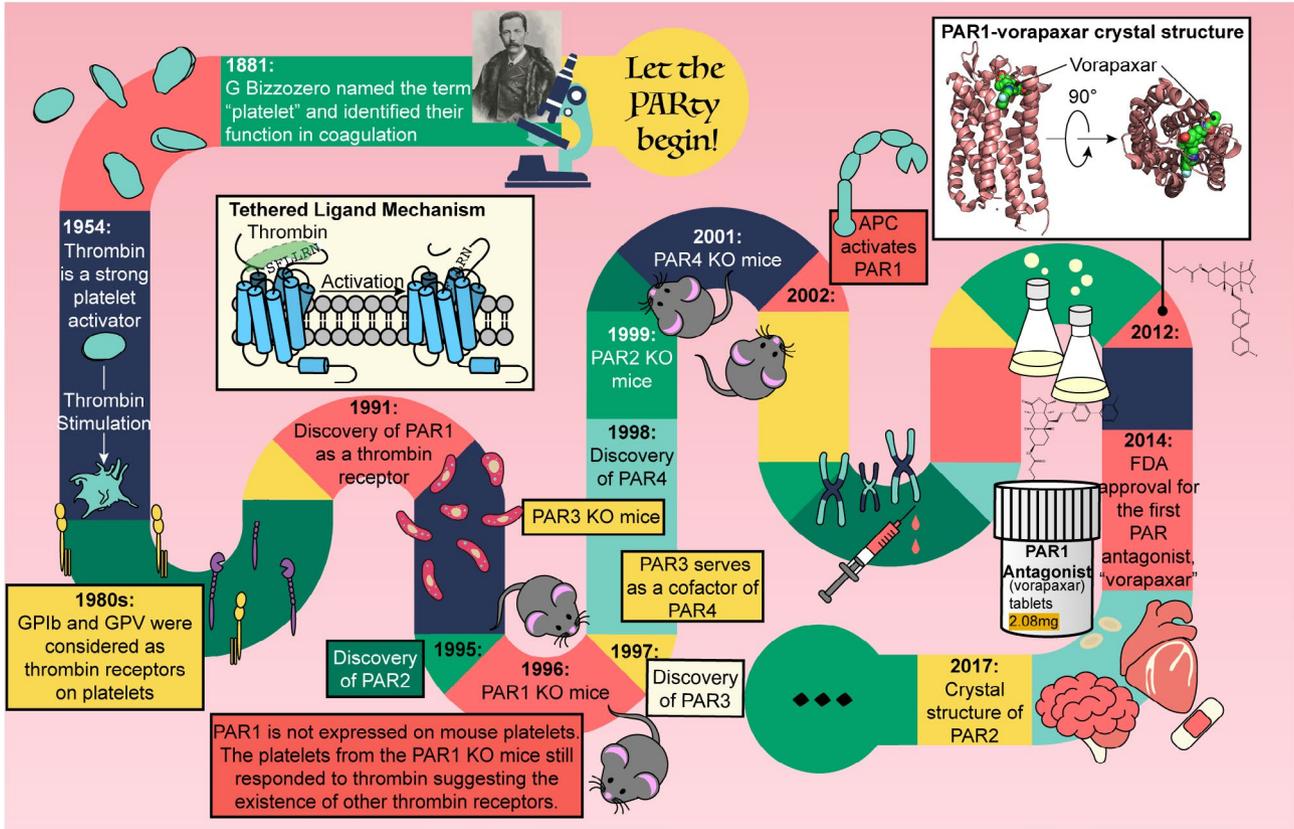
Essentials

- Protease-activated receptors (PARs) are G-protein-coupled receptors that mediate protease signaling.
- PARs are expressed widely in the body, and they can be activated by various proteases.
- Cofactors, cellular context, and the activating protease influence downstream signaling.
- PARs are promising therapeutic targets for antiplatelet and antithrombotic therapies.

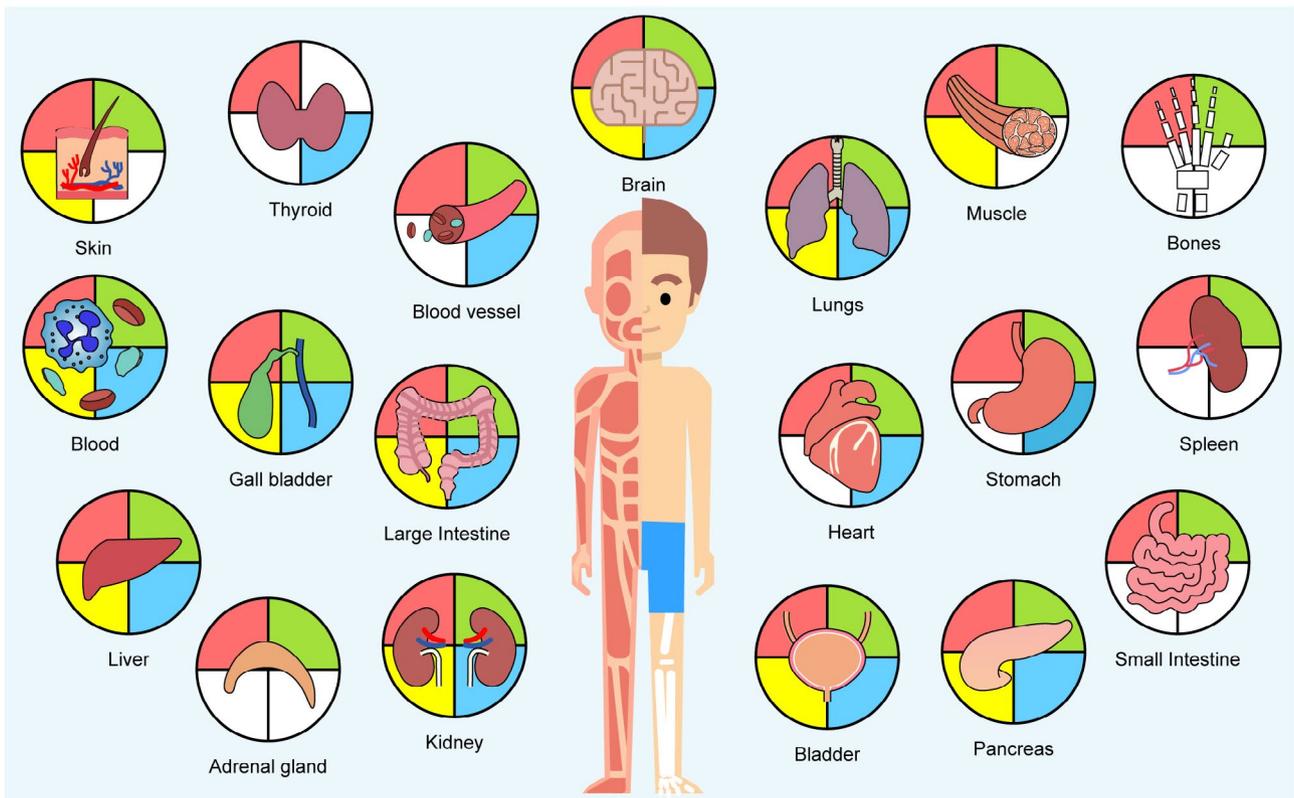
Abbreviations: ALIX, ALG2-interacting protein X; AP, adaptor protein; APC, activated protein C; ECL, extracellular loop; EPCR, endothelial protein C receptor; ESCRT, endosomal sorting complexes required for transport; GPCR, G-protein-coupled receptor; ICL, intracellular loop; MMP, matrix metalloprotease; PAR, protease-activated receptor; PC, protein C; TM, transmembrane.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2020 The Authors. *Research and Practice in Thrombosis and Haemostasis* published by Wiley Periodicals LLC on behalf of International Society on Thrombosis and Haemostasis (ISTH).

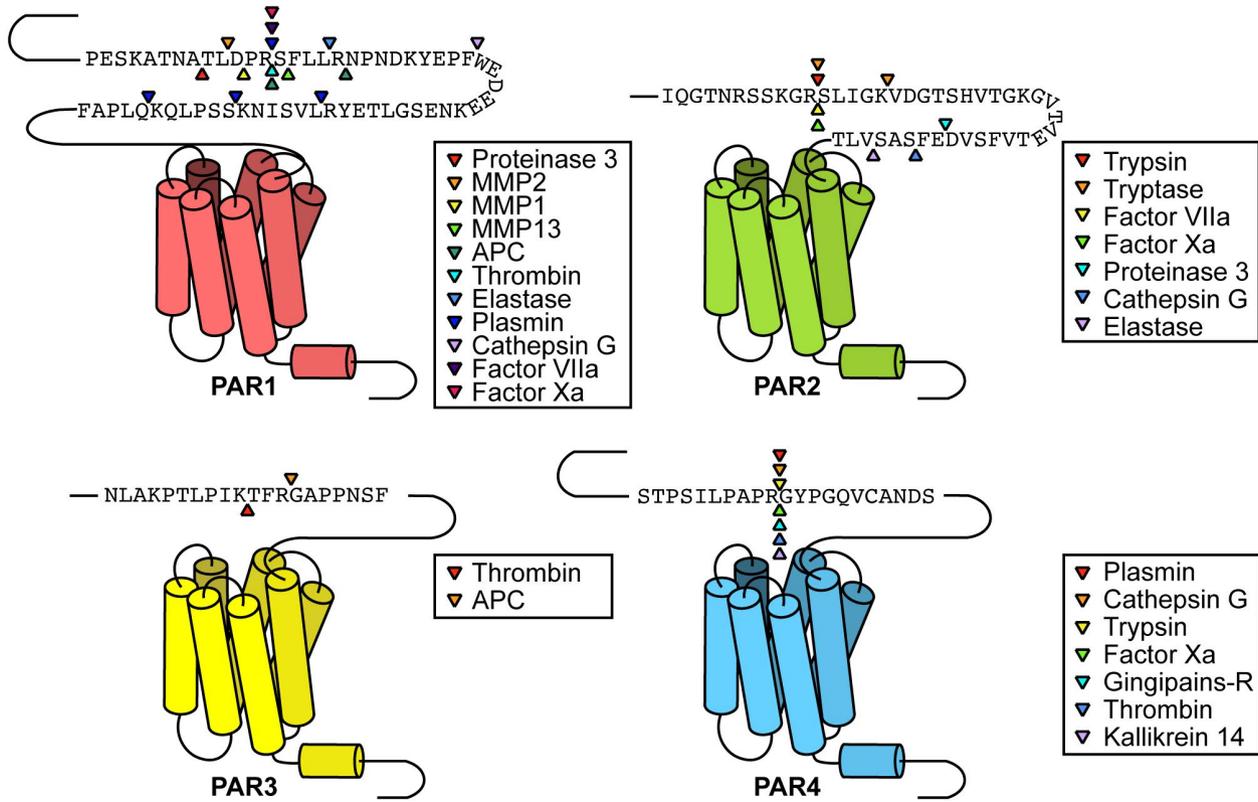


PAR Protein vs Gene Names: PAR1, F2R (Factor II Receptor); PAR2, F2RL1 (Factor II Receptor-Like 1); PAR3, F2RL2 (Factor II Receptor-Like 2); PAR4, F2RL3 (Factor II Receptor-Like 3) [1-3]



PAR expression as identified by expression analysis or experimental data:
PAR1: red; PAR2: green; PAR3: yellow; PAR4: blue; no known expression: white [4]

Proteases that cleave PARs and their respective cleavage sites [2, 5, 6]



Using platelets as an example, platelet PAR expression profiles vary by species. Human platelets have PAR1 and PAR4. However, mouse platelets express PAR3 and PAR4. Therefore, mice have limited utility as a preclinical model for developing PAR antagonists. Translational studies generally require experiments in non-human primates.

Jerry >

Hey man, can I get some cells from you for my PAR experiments?

Depends... Remember we have different PAR profiles!

Oh yeah... can I get some platelets from you to test my PAR1 inhibitor?

COME ON, MAN! I don't even have PAR1 on my platelets!

PAR Expression in Platelets from Different Species

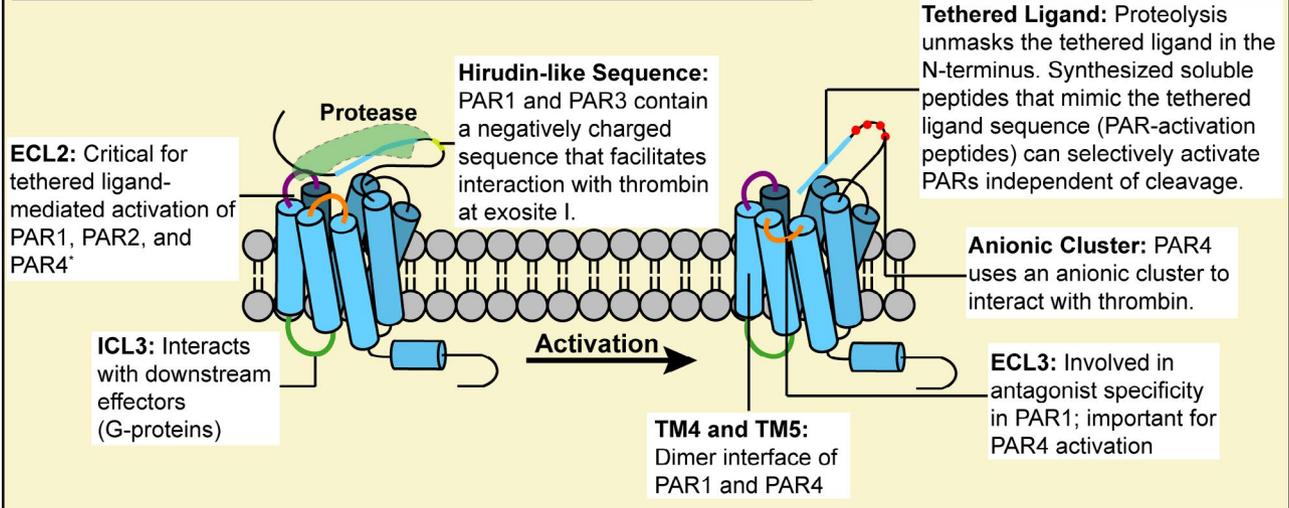
Species	PAR1	PAR3	PAR4
Human	X		X
Mouse		X	X
Rat		X	X
Rabbit		X	X
Guinea pig	X	X	X
Baboon			X
Monkey	X		X

You need to ask Abu for this!

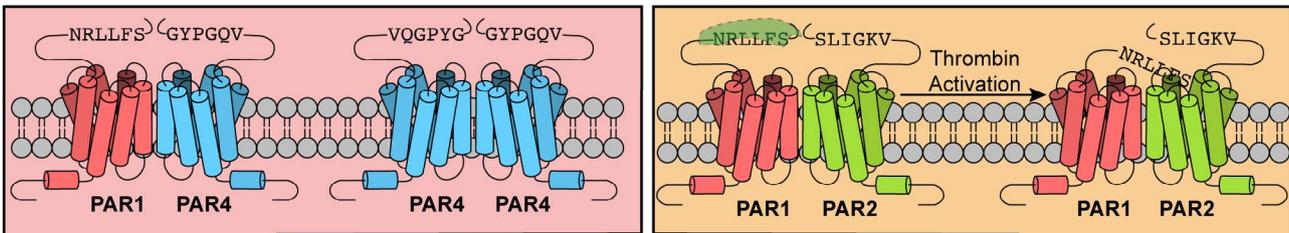
Expression of PARs across Species [2]

Animal models are essential for examining PAR function *in vivo*. However, the PAR expression profiles differ depending on cell type and species.

Tethered Ligand Mediated PAR Activation Mechanism [2, 5]



Note: Mouse PAR3 is not known to signal independently whereas independent signaling via human PAR3 remains controversial

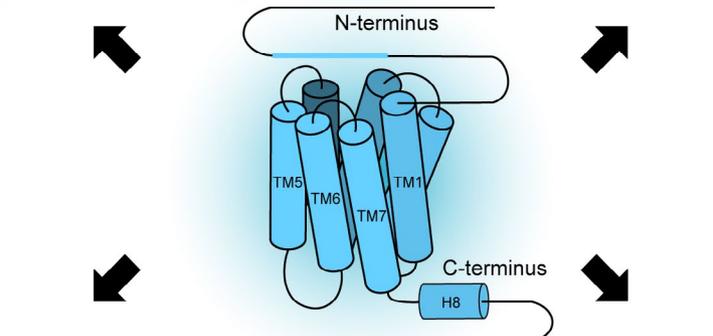


PARs form functional homo- and hetero-dimers with themselves or other PARs to regulate signaling. [2, 5]

Homo- and Hetero-Dimers

Transactivation

PAR1 and PAR2 form hetero-dimers to facilitate transactivation of PAR2 by thrombin-cleaved PAR1. [2, 7]

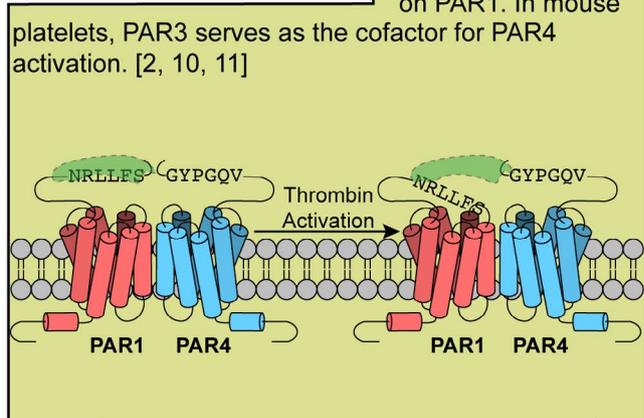
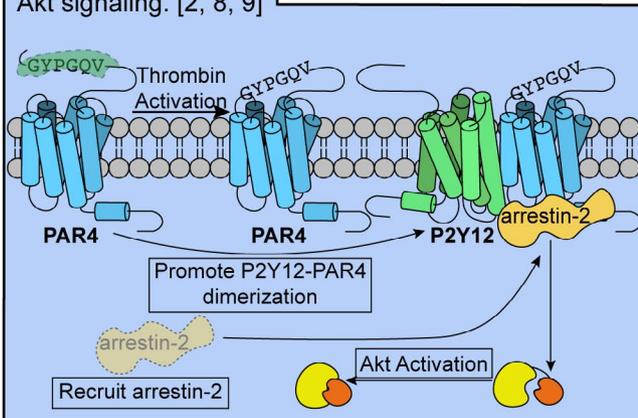


Allosteric Modulation

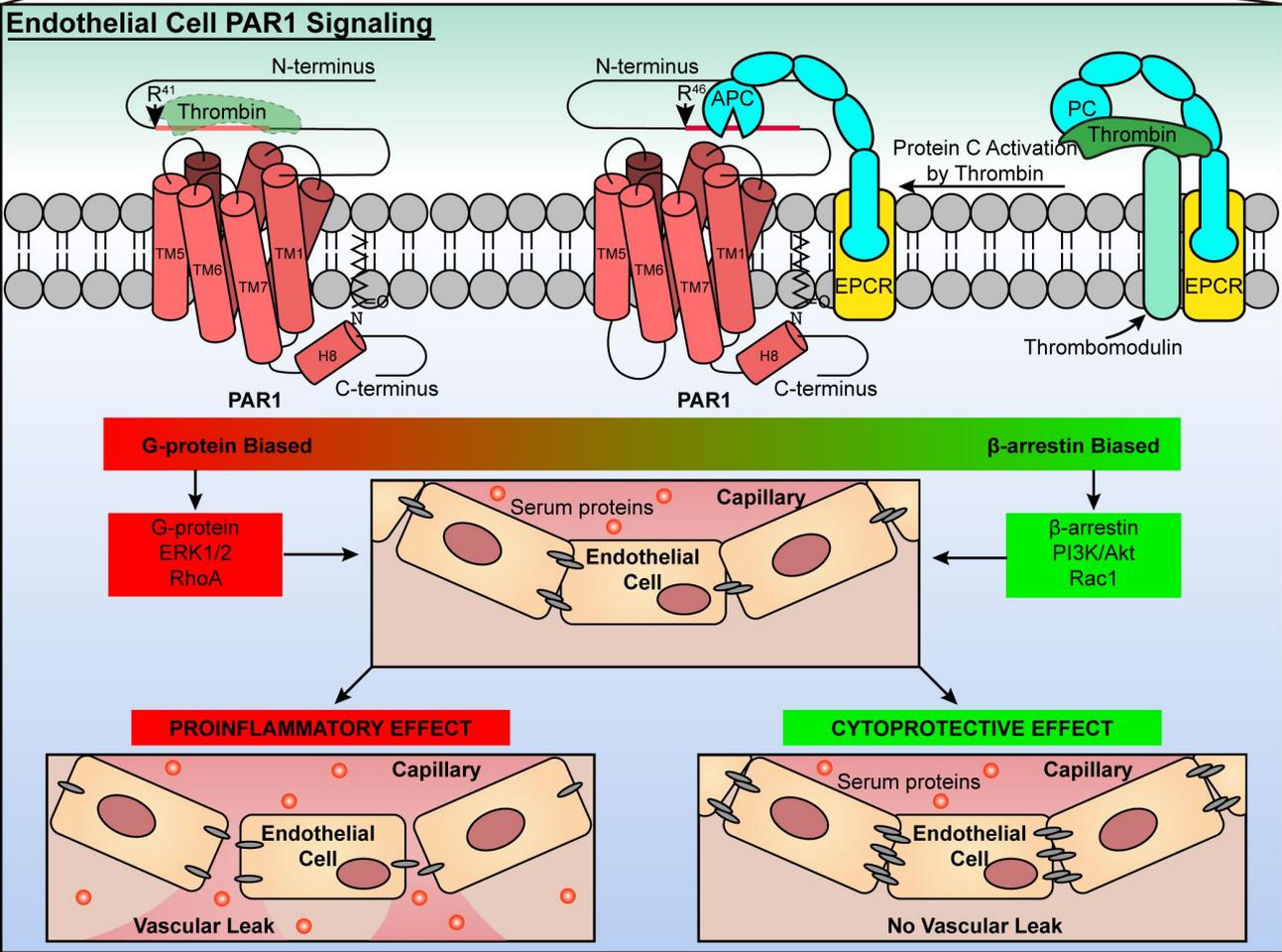
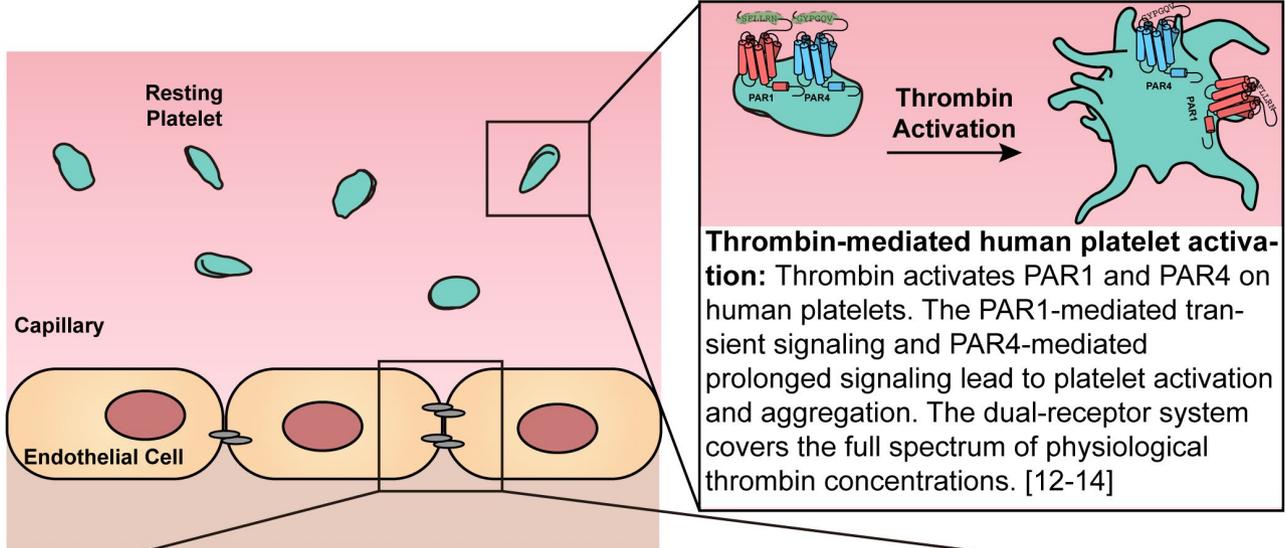
Cofactor Functions

Thrombin-mediated PAR4 activation promotes the formation of P2Y12-PAR4 hetero-dimer, enhances the recruitment of arrestin-2, and supports prolonged Akt signaling. [2, 8, 9]

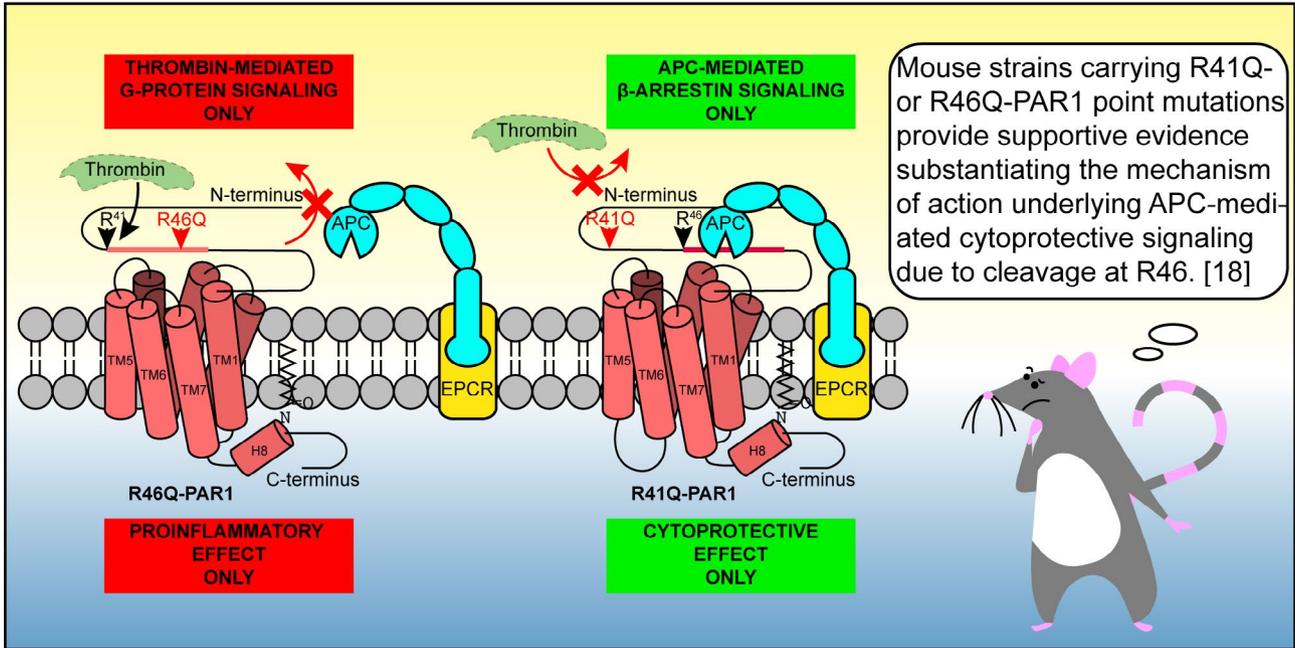
PAR1 serves as a cofactor for thrombin-mediated PAR4 activation. The enhanced rate of PAR4 activation by thrombin is facilitated by the hirudin-like sequence on PAR1. In mouse platelets, PAR3 serves as the cofactor for PAR4 activation. [2, 10, 11]



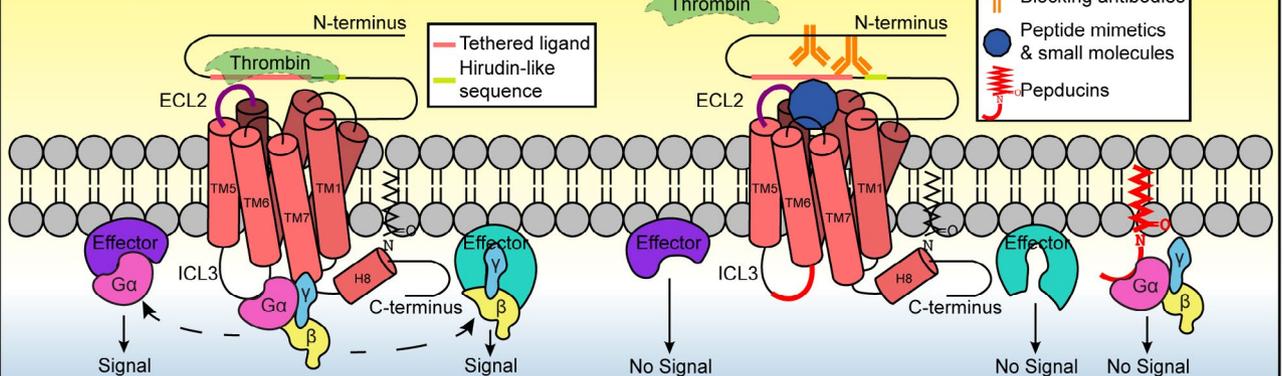
CELLULAR OUTCOMES OF PAR1 BIASED SIGNALING



Thrombin-mediated PAR1 activation leads to proinflammatory signaling and vascular leakage. Under physiological conditions, endothelial protein C receptor (EPCR) is a critical co-factor that facilitates APC-mediated PAR1 cleavage. In the lipid-raft/caveolae microenvironment, thrombin binds thrombomodulin and activates protein C that is bound to EPCR. This specific microenvironment allows PAR1 to be activated by EPCR-bound APC, but not thrombomodulin-bound thrombin. The EPCR-APC-mediated PAR1 activation triggers cytoprotective signaling and stabilizes the vessel wall. [15-17]

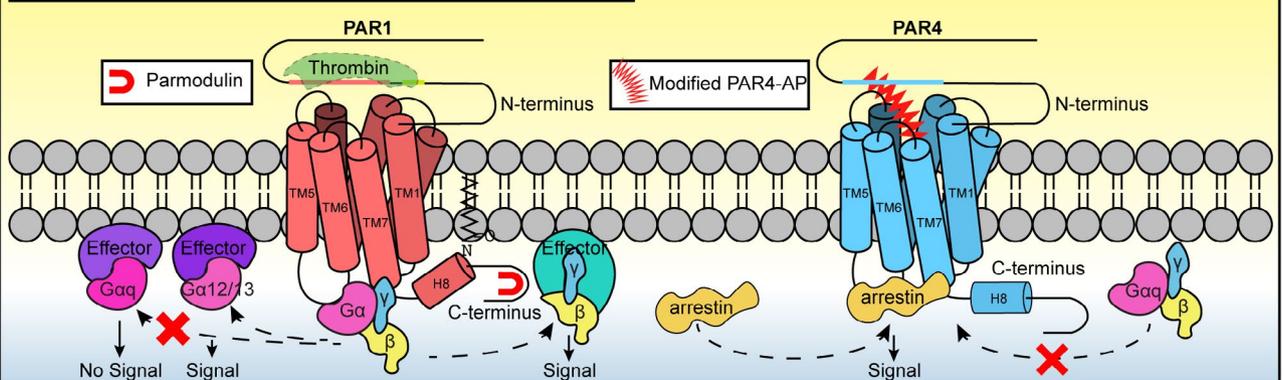


PAR Antagonist Mechanisms

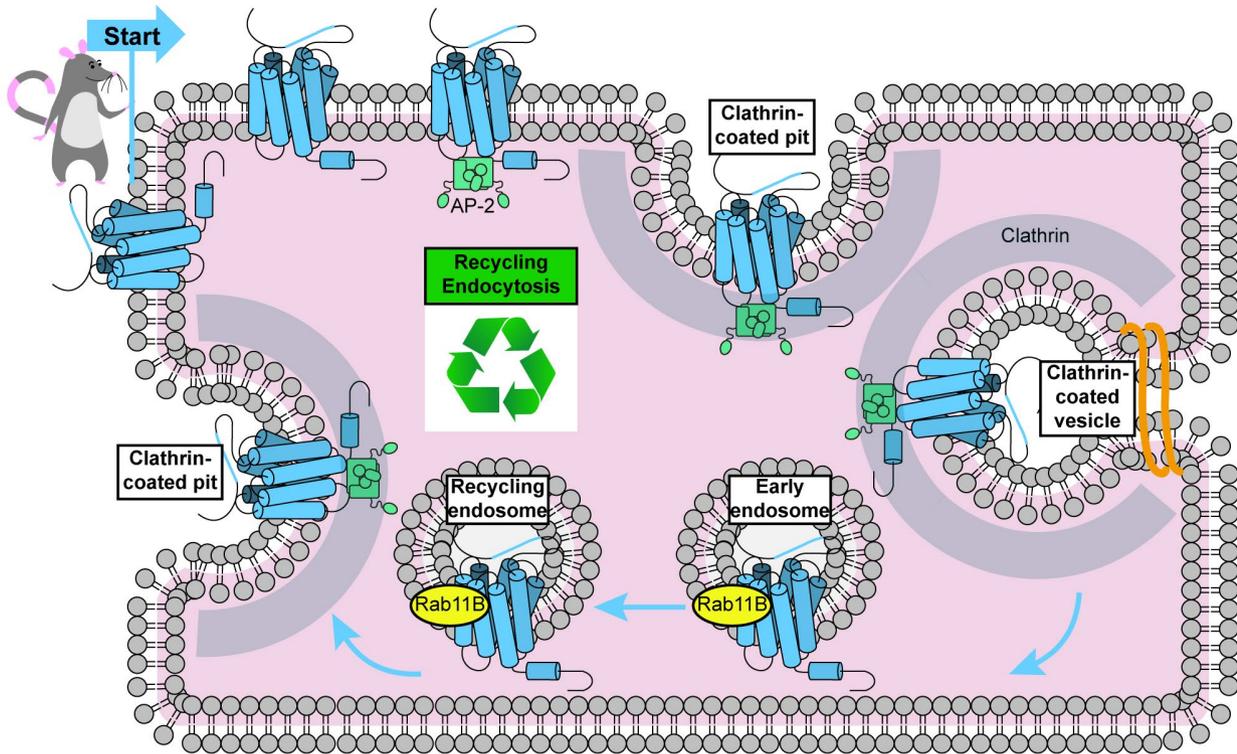


Using PAR1 as an example, there are 3 strategies for blocking PAR signaling. [5, 19] 1) Blocking antibodies prevent protease engagement and cleavage; 2) Peptide mimetics and small molecules block the tethered ligand binding site; and 3) Pepducins act as a decoy to sequester G-proteins away from the activated receptors.

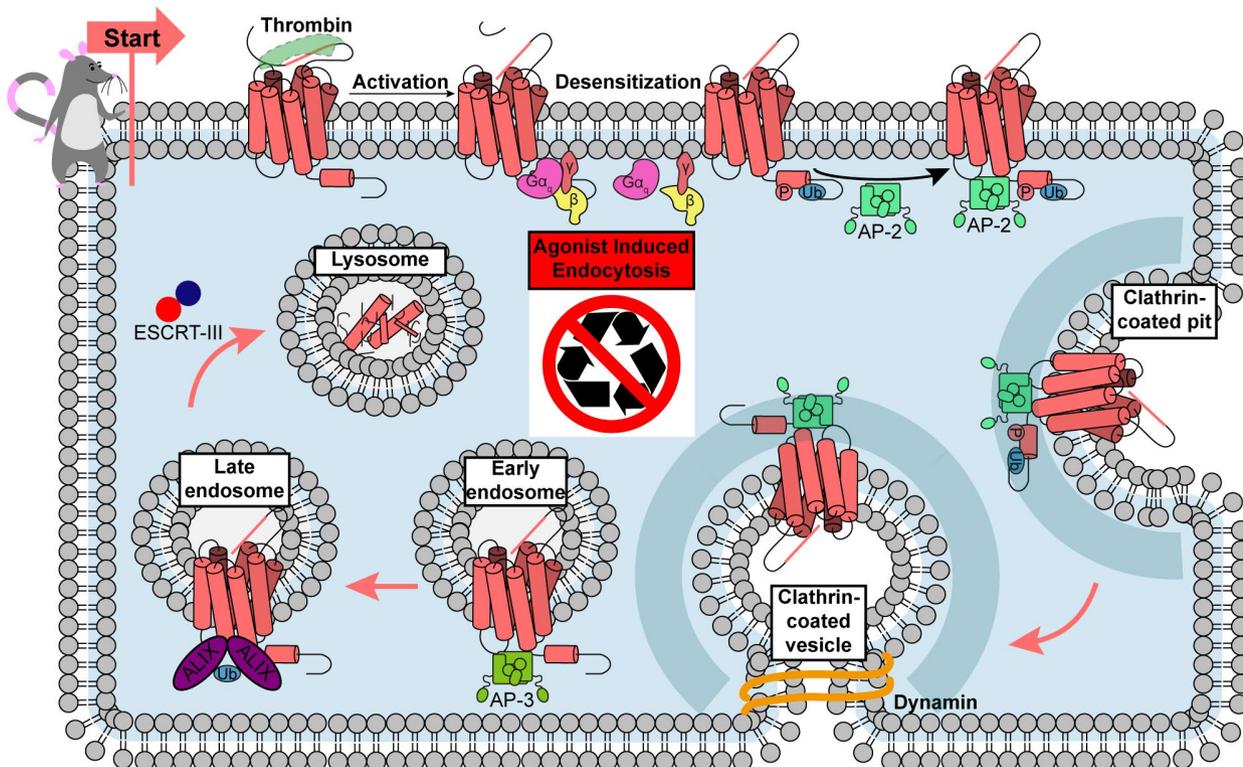
Pharmacological Redirection of PAR Signaling

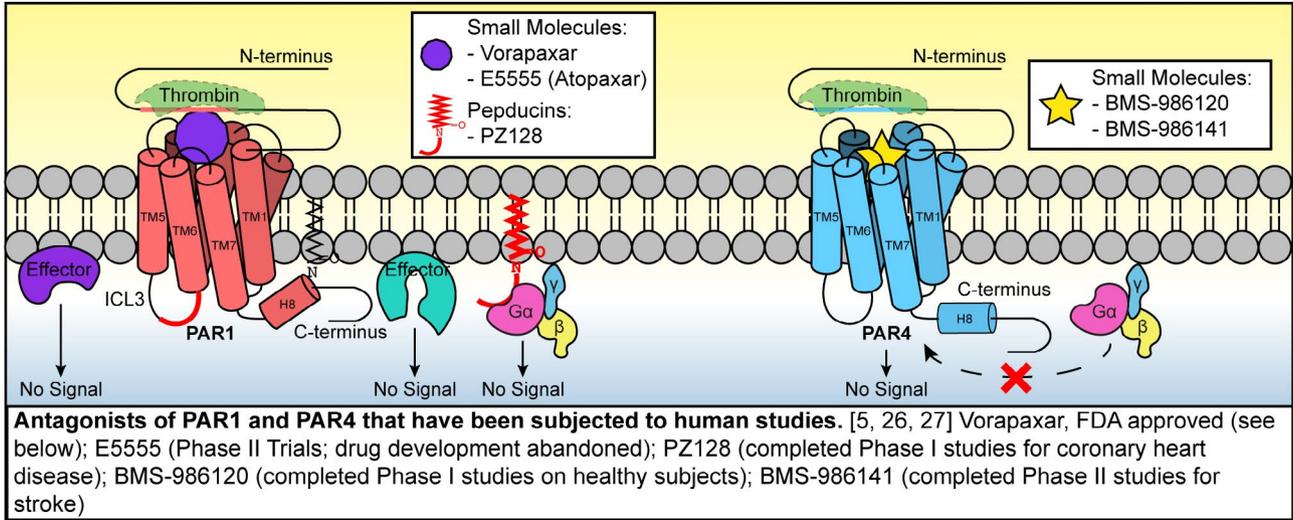


Two methods have been developed to guide downstream signaling outcomes, for example: 1) Parmodulin binds to the PAR1 C-terminus to preferentially engage and activate G $\alpha_{12/13}$ and 2) Modified PAR4-activation peptide (PAR4-AP) biases activated PAR4 towards arrestin engagement as opposed to G-protein signaling. [20-22]



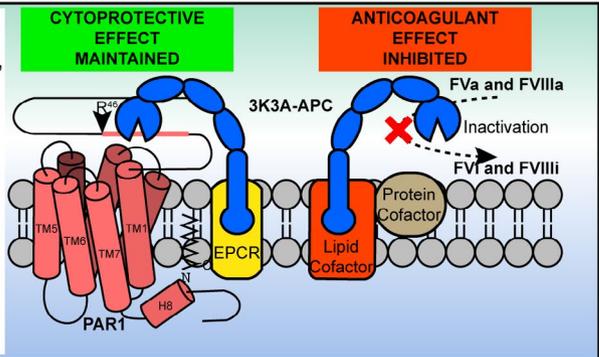
Recycling and Agonist-Induced Endocytic Degradation. Top panel: Constitutive recycling of naïve PARs. The internalization starts with AP-2 binding to PARs, and ends with the receptor recycling back to the cell surface in a Rab11B-dependent manner. Bottom panel: Because cleavage-mediated PAR activation is irreversible, the receptor must be desensitized and eventually degraded to permanently inactivate the signal. AP-2 recognizes activation-dependent phosphorylation and ubiquitination of PARs to initiate receptor internalization. AP-3, ALIX, ESCRT-III are important regulators of endosomal sorting to escort activated-PARs to lysosomes for degradation. [23-25]





Vorapaxar: Target: PAR1 [28, 29]
First in class PAR antagonist to receive FDA-approval (2014)
Indications:
- History of (1) Myocardial Infarction (MI) or (2) Established Peripheral Arterial Disease (PAD)
Contraindications and Black Box Warning:
- History of (1) Stroke, (2) Transient Ischemic Attack (TIA), (3) Intracranial Hemorrhage (ICH), or (4) Active Bleeding
- Antiplatelet agents, including vorapaxar, increase the risk of bleeding, including ICH and fatal bleeding.
Dose: 2.08 mg daily in combination with aspirin and/or clopidogrel.
Due to its very long half-life, vorapaxar is effectively an irreversible platelet antagonist.

Clinical Trials of 3K3A-APC [30, 31]
APC is a serine protease that has antithrombotic, antiinflammatory, antiapoptotic, and cytoprotective effects. Previous clinical studies of recombinant APC were abandoned due to increased bleeding risk. This molecularly engineered recombinant APC variant (3K3A-APC) retains its PAR1-mediated cytoprotective signaling function but retains <10% of its anticoagulant activity.
The NeuroNEXT consortium conducted a Phase II study of 3K3A-APC for ischemic stroke which was completed in 2018 and demonstrated neuroprotective effects with a trend toward reduced intraparenchymal hemorrhage.



PAR4 reactivity is affected by single nucleotide polymorphisms (SNPs) [5]

SNP	Residue	Alleles	Location	Effect	Potential Mechanism
rs773902*	120	Thr→Ala	TM2	↓ reactivity	Close to ligand binding site
	157	Tyr→Cys	TM3	↓ reactivity	Affects receptor transportation
rs2227346	296	Phe→Val	TM6	↓ reactivity	Affects ligand binding/Na ⁺ pocket microswitch
rs2227376	310	Pro→Leu	ECL3	↓ reactivity	Affects ECL3 rigidity

* The minor allele frequency (MAF) for the Thr¹²⁰ is 0.19 in individuals with European ancestry and 0.45 for those with African ancestry

PARting Conclusions: PARs have a broad tissue expression profile and flexible downstream signaling outcomes, suggesting important roles in (patho)-physiologies beyond hemostasis and vascular biology that may impact multiple organ systems. Thus PAR-directed pharmacotherapeutics may hold promise in variety of diseases, but the widespread tissue expression of PARs may lead to unintended toxicities that need to be considered in preclinical models and clinical trials.

ACKNOWLEDGMENTS

The authors regret that work from some colleagues could not be referenced or discussed due to space limitations, in particular, the experimental works identifying tissue expression of PARs. The authors would thank Maria de la Fuente, Elizabeth Knauss, and Amanda Waller for critical suggestions and helpful discussion.

RELATIONSHIP DISCLOSURE

The authors declare no conflicts of interest.

AUTHOR CONTRIBUTIONS

XH, MN, and BAK wrote the manuscript.

ORCID

Xu Han  <https://orcid.org/0000-0002-1977-1046>

Marvin T. Nieman  <https://orcid.org/0000-0003-2602-023X>

Bryce A. Kerlin  <https://orcid.org/0000-0002-1756-8271>

TWITTER

Marvin T. Nieman  @marvnieman

Bryce A. Kerlin  @drklotter

REFERENCES

- Coller BS. Historical perspective and future directions in platelet research. *J Thromb Haemost.* 2011;9(suppl 1):374–95.
- Han X, Bouck EG, Zunica ER, Arachiche A, Nieman M. Protease-activated receptors. In: Michelson DA, ed. *Platelets Fourth Edition*, 4th edn. Cambridge, MA: Academic Press; 2019: 243.
- Vu TK, Hung DT, Wheaton VI, Coughlin SR. Molecular cloning of a functional thrombin receptor reveals a novel proteolytic mechanism of receptor activation. *Cell.* 1991;64(6):1057–68.
- Petryszak R, Burdett T, Fiorelli B, Fonseca NA, Gonzalez-Porta M, Hastings E, et al. Expression Atlas update—a database of gene and transcript expression from microarray- and sequencing-based functional genomics experiments. *Nucleic Acids Res.* 2014;42:D926–32.
- Han X, Nieman M. The domino effect triggered by the tethered ligand of the protease activated receptors. *Thromb Res.* 2020;196:87–98.
- Madhusudhan T, Kerlin BA, Isermann B. The emerging role of coagulation proteases in kidney disease. *Nat Rev Nephrol.* 2016;12(2):94–109.
- Lin H, Trejo J. Transactivation of the PAR1-PAR2 heterodimer by thrombin elicits beta-arrestin-mediated endosomal signaling. *J Biol Chem.* 2013;288(16):11203–15.
- Khan A, Li D, Ibrahim S, Smyth E, Woulfe DS. The physical association of the P2Y12 receptor with PAR4 regulates arrestin-mediated Akt activation. *Mol Pharmacol.* 2014;86(1):1–11.
- Smith TH, Li JG, Dores MR, Trejo J. Protease-activated receptor-4 and purinergic receptor P2Y12 dimerize, co-internalize, and activate Akt signaling via endosomal recruitment of beta-arrestin. *J Biol Chem.* 2017;292(33):13867–78.
- Arachiche A, Mumaw MM, de la Fuente M, Nieman MT. Protease-activated receptor 1 (PAR1) and PAR4 heterodimers are required for PAR1-enhanced cleavage of PAR4 by alpha-thrombin. *J Biol Chem.* 2013;288(45):32553–62.
- Sambrano GR, Weiss EJ, Zheng YW, Huang W, Coughlin SR. Role of thrombin signalling in platelets in haemostasis and thrombosis. *Nature.* 2001;413(6851):74–8.
- Kahn ML, Nakanishi-Matsui M, Shapiro MJ, Ishihara H, Coughlin SR. Protease-activated receptors 1 and 4 mediate activation of human platelets by thrombin. *J Clin Invest.* 1999;103(6): 879–87.
- Kahn ML, Zheng YW, Huang W, Bigornia V, Zeng D, Moff S, et al. A dual thrombin receptor system for platelet activation. *Nature.* 1998;394(6694):690–4.
- Sveshnikova AN, Balatskiy AV, Demianova AS, Shepelyuk TO, Shakhidzhanov SS, Balatskaya MN, et al. Systems biology insights into the meaning of the platelet's dual-receptor thrombin signaling. *J Thromb Haemost.* 2016;14(10):2045–57.
- Komarova Y, Malik AB. Regulation of endothelial permeability via paracellular and transcellular transport pathways. *Annu Rev Physiol.* 2010;72:463–93.
- Feistritz C, Schuepbach RA, Mosnier LO, Bush LA, Di Cera E, Griffin JH, et al. Protective signaling by activated protein C is mechanistically linked to protein C activation on endothelial cells. *J Biol Chem.* 2006;281(29):20077–84.
- Zhao P, Metcalf M, Bunnett NW. Biased signaling of protease-activated receptors. *Front Endocrinol (Lausanne).* 2014;5:67.
- Sinha RK, Wang Y, Zhao Z, Xu X, Burnier L, Gupta N, et al. PAR1 biased signaling is required for activated protein C in vivo benefits in sepsis and stroke. *Blood.* 2018;131(11):1163–71.
- Hamilton JR, Trejo J. Challenges and opportunities in protease-activated receptor drug development. *Annu Rev Pharmacol Toxicol.* 2017;57:349–73.
- Aisiku O, Peters CG, De Ceunynck K, Ghosh CC, Dilks JR, Fustolo-Gunnink SF, et al. Parmodulins inhibit thrombus formation without inducing endothelial injury caused by vorapaxar. *Blood.* 2015;125(12):1976–85.
- De Ceunynck K, Peters CG, Jain A, Higgins SJ, Aisiku O, Fitch-Tewfik JL, et al. PAR1 agonists stimulate APC-like endothelial cytoprotection and confer resistance to thromboinflammatory injury. *Proc Natl Acad Sci U S A.* 2018;115(5):E982–E91.
- Thibeault PE, LeSarge JC, Arends D, Fernandes M, Chidiac P, Stathopoulos PB, et al. Molecular basis for activation and biased signaling at the thrombin-activated GPCR proteinase activated receptor-4 (PAR4). *J Biol Chem.* 2020;295(8):2520–40.
- Arakaki AKS, Pan WA, Trejo J. GPCRs in cancer: protease-activated receptors, endocytic adaptors and signaling. *Int J Mol Sci.* 2018;19(7).
- Marchese A, Paing MM, Temple BR, Trejo J. G protein-coupled receptor sorting to endosomes and lysosomes. *Annu Rev Pharmacol Toxicol.* 2008;48:601–29.
- Grimsey N, Lin H, Trejo J. Endosomal signaling by protease-activated receptors. *Methods Enzymol.* 2014;535:389–401.
- Sidhu TS, French SL, Hamilton JR. Differential signaling by protease-activated receptors: implications for therapeutic targeting. *Int J Mol Sci.* 2014;15(4):6169–83.
- Gurbel PA, Bliden KP, Turner SE, Tantry US, Gesheff MG, Barr TP, et al. Cell-Penetrating Pepducin Therapy Targeting PAR1 in Subjects With Coronary Artery Disease. *Arterioscler Thromb Vasc Biol.* 2016;36(1):189–97.
- Frampton JE. Vorapaxar: a review of its use in the long-term secondary prevention of atherothrombotic events. *Drugs.* 2015;75(7):797–808.
- Gryka RJ, Buckley LF, Anderson SM. Vorapaxar: the current role and future directions of a novel protease-activated receptor antagonist for risk reduction in atherosclerotic disease. *Drugs R D.* 2017;17(1):65–72.
- Lyden P, Pryor KE, Coffey CS, Cudkowicz M, Conwit R, Jadhav A, et al. Final results of the RHAPSODY trial: a multi-center, phase 2 trial using a continual reassessment method to determine the safety

and tolerability of 3K3A-APC, a recombinant variant of human activated protein C, in combination with tissue plasminogen activator, mechanical thrombectomy or both in moderate to severe acute ischemic stroke. *Ann Neurol.* 2019;85(1):125–36.

31. Griffin JH, Fernandez JA, Lyden PD, Zlokovic BV. Activated protein C promotes neuroprotection: mechanisms and translation to the clinic. *Thromb Res.* 2016;141(suppl 2):S62–4.

How to cite this article: Han X, Nieman MT, Kerlin BA. Protease-activated receptors: An illustrated review. *Res Pract Thromb Haemost.* 2021;5:17–26. <https://doi.org/10.1002/rth2.12454>